Mast Group Ltd.
Mast House, Derby Road,
Bootle, Merseyside, L20 1EA
United Kingdom
Tel: + 44 (0) 151 472 1444
Fax: + 44 (0) 151 944 1332
email: sales@mast-group.com
Web: www.mast-group.com

Mast Diagnostica GmbH
Feldstrasse 20
DE-23858 Reinfeld
Germany
Tel: + 49 (0) 4533 2007 0
Fax: + 49 (0) 4533 2007 68
email: mast@mast-diagnostica.de
Web: www.mast-group.com

Mast Diagnostic
12 rue Jean-Jacques Mention
CS91106, 80011 Amiens, CEDEX 1
France
Tél: + 33 (0) 3 22 80 80 67
Fax: + 33 (0) 3 22 80 99 22
email: info@mast-diagnostic.fr
Web: www.mast-group.com

Chloramphenicol
MAST® SELECTAVIAL

SV54 Series

Intended use
For the isolation of yeasts and moulds from food, environmental and clinical specimens.

FOR IN VITRO DIAGNOSTIC USE ONLY

Contents
10 vials of MAST® SELECTAVIAL.

Formulation

Material:	Concentration in medium:
Chloramphenicol	100 mg/L

Storage and shelf life
Store unopened at 2 to 8°C until expiry date shown on pack label. Once reconstituted use immediately.

Precautions
For *in vitro* diagnostic use only. Observe approved biohazard precautions and aseptic techniques. To be used only by adequately trained and qualified laboratory personnel. Sterilise all biohazard waste before disposal. Refer to Product Safety Data sheet.

Materials required but not provided
Standard microbiological supplies and equipment such as loops, MAST culture media, swabs, applicator sticks, incinerators and incubators, etc., as well as serological and biochemical reagents and additives such as blood.

Procedure
1. Sterilise the appropriate volume of MAST® Sabouraud Dextrose Agar (DM200D), cool to 50 to 55°C and hold at this temperature.
2. Reconstitute the contents of one vial using the diluent specified on the pack label. The best method is to aseptically add the diluent using a sterile needle and syringe. Draw the diluent into the syringe and after removing the plastic cap, inject through the rubber stopper of the vial. The lyophilised supplement will rapidly dissolve and may be withdrawn into the syringe.
3. Add the antibiotic supplement to the volume of medium specified on the pack label and discard the needle into an approved container.
4. Mix gently but thoroughly to evenly distribute the selective agents. Pour culture plates (15 to 20 mL per plate) and allow to set.
5. Prepared culture plates may be used immediately or stored in plastic bags at 2 to 8°C for up to one week before use.

For clinical specimens:
Skin scales, nail clippings and scalp scrapings are the most suitable specimens for culture from sites of superficial infection. These can be directly inoculated into the medium. For subcutaneous mycoses, aspirated pus, tissue biopsies and scrapings of crusts are normally taken for culture and can be spread onto the surface of the medium. Plates should be incubated at 25 to 30°C for at least 2 to 3 weeks.

For food specimens:
Using a suitable diluent, prepare a 10^{-1} homogenate of the food to be tested and further serial dilutions as required. Use the homogenate and dilutions to inoculate a medium suitable for surface counting and incubate at 25 to 30°C for 3 to 5 days, then for up to 2 weeks to count very slow growing species. Count the number of yeast and mould colonies formed and report as the number per gram of original sample.

Interpretation of results
The following table shows examples of expected growth characteristics for yeasts and moulds:

Organism	Colony size (mm)	Colour	Other
C. albicans	0.5-2.0	White	Yeasty smell
C. krusei	1.0-3.0	Grey-white	Yeasty smell
M. canis	25	Reverse yellow	-

Quality control
Check for signs of deterioration. Quality control must be performed with at least one organism to demonstrate a positive reaction and at least one organism to demonstrate a negative reaction. Do not use the product if the reactions with the control organisms are incorrect. The list below illustrates a range of performance control strains which the end user can easily obtain.

Test Organisms	Result
Staphylococcus aureus ATCC® 25923	No growth
Candida albicans ATCC® 10231	Growth

References
Bibliography available on request.

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