

MAST® ID CHROMagar® *Candida*

IDM40

Intended use

For the simultaneous detection and presumptive identification of *Candida albicans*, *Candida tropicalis* and other *Candida* spp. and yeasts.

Contents

See pack label.

Formulation*

| Material: | Concentration in medium: |
|-------------------------|--------------------------|
| Peptone mixture | 10.0g/litre |
| Special chromogenic mix | 22.0g/litre |
| Chloramphenicol | 0.5g/litre |
| Agar | 15.0g/litre |
| Final pH: 6.3 ± 0.2 | |

Storage and shelf life

All dehydrated culture media containers should be kept tightly closed and stored in a dry place at 10 to 25°C until the expiry date shown on the pack label.

Precautions

For *in vitro* diagnostic use only. Observe approved hazard precautions and aseptic techniques. To be used only by adequately trained and qualified laboratory personnel. Sterilise all biohazard waste before disposal. Refer to Product Safety Data sheet (available on request or via MAST® website).

Materials required but not provided

Standard microbiological supplies and equipment such as loops, MAST® selective supplements, swabs, applicator sticks, incinerators and incubators, etc., as well as serological and biochemical reagents and additives such as blood.

Procedure

1. Refer to pack label for quantities and volumes required. Prepare MAST® IDCHROMagar® *Candida* (IDM40/A) by suspending the powder in distilled or deionised water. For sachet packs, dissolve the entire contents of the sachet in the volume shown on the label.
2. Bring to the boil and stir regularly until the agar is completely dissolved.
DO NOT AUTOCLAVE OR OVERHEAT.
3. Allow to cool to 50 to 55°C. Swirl to homogenise and pour culture plates (15 to 20ml per plate). Allow to set on a flat surface.
4. Prepared culture plates may be used immediately or stored in plastic bags at 2 to 8°C for up to one week.

5. Ensure the surface of the plate is dry before use.
6. Inoculate suspect colonies directly by surface plating, streaking out for single colonies.
7. Incubate plates aerobically at 35 to 37°C for 24 to 48 hours. Optimum colour intensity is achieved after 48 hours incubation.

Interpretation of results

After incubation record growth of organisms. Identify individual organisms by the colouring acquired during growth as well as colonial morphology.

| Colony colour | Microorganism |
|---------------|-------------------------------------|
| Green | Presumptive of <i>C. albicans</i> |
| Blue | Presumptive of <i>C. tropicalis</i> |
| White to Pink | Other species |

C.krusei forms pale pink to purple crenated, rough spreading colonies with pale edges.

T.beigelii forms pale "dirty pink" to "dirty green-grey" small colonies which become darker and rough on prolonged incubation (i.e. 72 hours).

Quality control

Check for signs of deterioration. Quality control must be performed with at least one organism to demonstrate expected performance. Do not use the product if the result with the control organism is incorrect. The list below illustrates a range of performance control strains which the end user can easily obtain.

| Test Organisms | Colony colour |
|--|----------------|
| <i>Candida albicans</i> ATCC® 90028 | Green |
| <i>Candida tropicalis</i> ATCC® 9968 | Blue |
| <i>Candida glabrata</i> ATCC® 90030 | Lilac |
| <i>Candida parapsilosis</i> ATCC® 90018 | Off white |
| <i>Candida krusei</i> ATCC® 6258 | Pink, crenated |

References

Bibliography available on request.