Mast Group Ltd.
Mast House, Derby Road,
Bootle, Merseyside, L20 1EA
United Kingdom
Tel: + 44 (0) 151 472 1444
Fax: + 44 (0) 151 944 1332
email: sales@mast-group.com
Web: www.mast-group.com

Mast Diagnostica GmbH
Feldstrasse 20
DE-23858 Reinfeld
Germany
Tel: + 49 (0) 4533 2007 0
Fax: + 49 (0) 4533 2007 68
email: mast@mast-diagnostica.de
Web: www.mast-group.com

Mast Diagnostic
12 rue Jean-Jacques Mention
CS91106, 80011 Amiens, CEDEX 1
France
Tél: + 33 (0) 3 22 80 80 67
Fax: + 33 (0) 3 22 80 99 22
email: info@mast-diagnostic.fr
Web: www.mast-group.com

Kimmig Agar Base

DM146

Intended Use
For the isolation, identification and cultivation of fungi.

Contents
See pack label.

Formulation*

Material:	Concentration in medium:
Peptone Mixture	13.0 g/litre
Sodium chloride	11.5 g/litre
D-glucose	10.0 g/litre
Agar	13.0 g/litre
Final pH: 6.5 ± 0.2	

Storage and shelf life
All dehydrated culture media containers should be kept tightly closed and stored in a dry place at 10 to 25°C until the expiry date shown on the pack label.

Precautions
For *in vitro* diagnostic use only. Observe approved hazard precautions and aseptic techniques. To be used only by adequately trained and qualified laboratory personnel. Sterilise all biohazard waste before disposal. Refer to Product Safety Data sheet (available on request or via MAST® website).

Materials required but not provided
Standard microbiological supplies and equipment such as loops, MAST® selective supplements, swabs, applicator sticks, incinerators and incubators, etc., as well as serological and biochemical reagents and additives such as blood.

Procedure
1. Refer to pack label for quantities and volumes required. Prepare MAST® Kimmig Agar (DM146D) by suspending the powder in distilled or deionised water. For sachet packs, dissolve the entire contents of the sachet in the volume shown on the label.
2. Add 5ml of glycerol to each litre of medium.
3. Autoclave at 121°C (15 p.s.i.) for 15 minutes.
4. Cool to 50 to 55°C and, if required for selective procedures, aseptically add antibiotics according to the methodology followed. Cycloheximide at a final concentration of 400mg/L, combined with either penicillin (40000 IU/L) and streptomycin (40mg/L), or colistin (80mg/L) and novobiocin (100mg/L) are commonly used.
5. Mix well, pour culture plates (15 to 20ml per plate) and allow to set.
6. Prepared culture plates may be used immediately or stored in plastic bags at 2 to 8°C for up to one week before use.
7. Inoculate plates with clinical, veterinary or food samples by surface plating, streaking out for single colonies.
8. Incubate plates aerobically for up to 3 weeks at 25 to 30°C.

Interpretation of results
After incubation record growth of organisms. Typical characteristics to note include: colony size, colour and morphology.

Quality control
Check for signs of deterioration. Quality control must be performed with at least one organism to demonstrate a positive reaction and at least one organism to demonstrate a negative reaction. Do not use the product if the reactions with the control organisms are incorrect. The list below illustrates a range of performance control strains which the end user can easily obtain.

Test Organisms	Result
Aspergillus niger ATCC® 16404	Growth, white/yellow mycelium with black sporing heads
Candida albicans ATCC® 90028	Growth, white colonies*
Candida krusei ATCC® 14243	Growth, white-grey colonies*

* On non-selective medium

References
Bibliography available on request.

IFU331 GB 08/20 V5

*Formulation may change to meet performance criteria